

HIV infection and bone changes

MD PHD Maria Carla Re

Section of Microbiology of the Department of Hematology, Oncologic Science, Anatomical Pathology and Microbiology and Interuniversity Consortium, National Institute Biostructure and Biosystem (INBB), Rome, Italy,

via Massarenti 11, 40138 Bologna (Bo) Italy

Tel.: +39 051 6361111 - Fax: +39 051 6361111 - E-mail: mariacarla.re@unibo.it

Key words: HIV; HAART; Bone changes

ABSTRACT

The introduction of highly active antiretroviral therapy (HAART) has radically changed the natural history of HIV infection. The significant reduction of plasma viremia accompanied by the arrested immune deterioration following administration of new therapeutic protocols between 1996 and 1999 generated cautious optimism also based on mathematical models claiming the possibility of a complete eradication of HIV infection. Even if the advent of HAART has significantly extended the life-span of patients living with AIDS, the increased mean life-span has given rise to long-term complications of HIV such as metabolic complication, cardiovascular disease and osteoporosis. Degenerative processes involving the bone compartment are common during the course of HIV-1 infection. Osteopenia and osteoporosis represent a challenge in clinical and therapeutic management. It has yet to be determined whether the cause of this dysfunction is linked to HIV-1-mediated direct and/or indirect effects on osteoblasts/osteoclasts cross-talk regulation. This report analyzes the numerous factors able to favor bone derangement (bone loss and osteopenia/osteoporosis) in HIV-1 infected patients.

Bone and bone changes

Bone is a specialized connective tissue characterized by hardness and plasticity, consisting of cells immersed in a mineralized extracellular matrix. Osteoblasts, characterized by the ability to secrete bone matrix (Harada & Rodan, 2003), and osteoclasts, responsible for bone resorption (Melissa et al., 2004), are involved in bone tissue regulation.

Both osteoblasts and osteoclasts act on bone homeostasis through a continuous remodelling process, necessary both to maintain calcium and phosphates homeostasis and to promote body adaptation to external tensional forces. The bone remodelling process begins with the proliferation and activation of osteoclasts on the bone surface and is controlled by systemic hormones like calcitonin, parathormone or estrogens, and local factors, most of them involved in inflammatory responses such as IL-1, IL-6, TNF- α and prostaglandins.

Moreover, the OPG/RANKL/RANK triad plays a central role in bone remodelling. In particular, OPG, released by osteoblasts, is a soluble RANKL receptor inhibiting the RANKL/RANK interaction and then the RANKL/RANK-mediated differentiation and activation of osteoclasts (Boyce & Xing, 2007). Remodelling rates of bone tissue can be monitored measuring molecular markers of bone remodelling. In particular, serum osteocalcin and alkaline phosphatase activity reflect bone formation, whereas products of catabolism

of the matrix, like cross-linked N-telopeptides of type I collagen (NTx) or C-telopeptides, are indicative of rates of bone resorption (Camozzi et al., 2007).

Osteoporosis represents severe derangement of bone tissue and is considered a skeletal disease characterized by low bone mass and bone micro-architectural deterioration with a major increase in bone fragility and fracture susceptibility.

Clinically osteoporosis is often not disclosed until the occurrence of low-trauma fractures, defined as fragility fractures (Becker, 2006). Fragility fractures mainly involve the vertebral body, distal forearm or hip and have significant effects on patients' quality of life with serious disability and excess mortality in the older subjects. This severe clinical evolution coupled with the high incidence of osteoporosis in the population indicate a central role of this disease in public health.

The progressive bone mass loss is classified by WHO as osteopenia or osteoporosis by dual-energy x-ray absorptiometry (DXA) assay which determines bone mineral density (BMD). DXA results are expressed in terms of T-score, that is the number of standard deviations below or above the mean of BMD of a population matched for race and sex (Cummings et al., 2002). Osteoporosis is characterized by a T-score value lower than -2.5, whereas a T-score between -2.5 and -1 indicates osteopenia and normality is defined as a T-score above -1 (Johnell et al., 2005).

The role of HIV-1 infection

The bone mass loss pathogenesis during HIV infection remains unsettled. Early studies on HIV/osteoblasts interaction to determine whether HIV is able to infect osteoblasts, have yielded controversial results (Mellert *et al.*, 1990; Campbell *et al.*, 1996; Nacher *et al.*, 2001). Several HIV proteins may affect the functionality and maturation of osteoblasts inducing apoptotic stimuli in mesenchymal stem cells, which are precursors of osteoblasts (Wang *et al.*, 2002). In particular HIV-1-gp120 and p55gag can reduce bone alkaline phosphatase activity and calcium deposition by osteoblasts, and Rev and p55gag, but not gp120, affect the differentiation of mesenchymal stem cells toward osteoblastic lineage (Cotter *et al.*, 2007).

In addition we recently demonstrated (Gibellini *et al.* 2008) that HIV-1, heat-inactivated HIV-1 and recombinant gp120 trigger apoptosis in primary osteoblasts and HOBIT cells, suggesting that HIV-1 does not infect osteoblasts, and that apoptosis induction is related to gp120/cell membrane interaction, resembling a mechanism similar to that already demonstrated in the derangement of haematopoietic CD34+ progenitor cells (Re *et al.*, 1994, Gibellini *et al.* 2007a,b, De Crignis *et al.* 2008).

The most important regulative mechanism of osteoclast/osteoblast activity is the OPG/RANKL/RANK system (Boyce & Xing, 2007). RANKL and OPG are yielded by lymphocytes (Chakravarti *et al.*, 2007; Ly *et al.*, 2007) other than osteoblasts, and increased levels of these cytokines are common in diseases characterized by persistent immune activation (Ueland *et al.*, 2001) that can determine bone loss (Clowes *et al.*, 2005, Seminari *et al.*, 2005; Gibellini *et al.*, 2007; Mora *et al.*, 2007). Interestingly, RANKL can be also up-regulated by viral proteins such as gp120 and Vpr (Fakruddin & Laurence, 2004, 2005) suggesting an osteoclastic hyper-activation in the bone compartment with bone homeostasis imbalance. Osteoclasts can also differentiate under the influence of TNF- α , (Breen, 2002) that may play an etiological role in bone loss for TNF- α (Aukrust *et al.*, 1999).

In addition to the direct role of HIV on osteoblast and osteoclast interaction, HIV has been associated with some risk factors for osteoporosis (Tomazic *et al.*, 2007) such as undernutrition and malabsorption pathologies which can influence levels of vitamin D and calcium (Cashman, 2007), or endocrine complications, such as androgen and estrogen deficiency (Madeddu *et al.*, 2004).

In the HIV seronegative population osteopenia and osteoporosis are mostly observed among women (Bonnick, 2006) where bone metabolism is related to hormonal status because of the protective role of estrogens, which mainly consists in down-regulating the production of pro-inflammatory cytokines as IL-1, IL6 and RANKL thereby inhibiting osteoclast differentiation (Sorensen *et al.*, 2006).

The extended course of viral infection and prolonged HAART treatment have displayed several degenerative aspects of HIV-related disease

regarding different cell lineages. In particular, HIV infection *per se* and HAART therapy can directly affect the homeostasis of bone tissue. Even if antiretroviral therapy may play an important role in bone loss, a significant decrease of bone density has been observed in HIV positive naïve patients (Loiseau-Peres *et al.*, 2002; Annapoorna *et al.*, 2004; Amorosa & Tebas 2006). Several data demonstrated a correlation between HIV-1 infection and bone loss. A marked decrease in bone turnover and reduced bone formation, more evident in patients with advanced progression of HIV disease (Serrano *et al.*, 1995, Thomas & Doherty, 2003; Bruera *et al.*, 2003; Amiel *et al.*, 2004) and a significant decrease of osteocalcin serum levels and increase in matrix degradation products are reported in HIV-positive individuals (Teichmann *et al.*, 2000).

In addition, the HAART/bone interaction has been studied to determine whether multidrug treatment is involved in BMD decrease (Carr *et al.*, 2001; Bongiovanni *et al.*, 2005; Vescini *et al.*, 2005; Aparicio *et al.*, 2006). Despite a high variability related to specific drug class and treatment duration, HAART seems to have a pivotal role, especially when including a protease inhibitor (PI) (Tebas *et al.*, 2000, Jain & Lenhard, 2002).

Conversely, other reports failed to reveal any influence of HAART on bone condition, finding no differences in BMD reduction between naïve and treated patients (Bruera *et al.*, 2003; Dube *et al.*, 2002; Landonio *et al.*, 2004; Amiel *et al.*, 2004). A dual HAART-induced effect has been hypothesized. The drugs initially exacerbate abnormalities in bone homeostasis (Mallon *et al.*, 2003) and then the restoration of some cytokine networks allows the normalization of bone remodelling process. A recent study performed on a large number of HIV-1 infected patients reported a high percentage of osteopenia and osteoporosis suggesting a direct role of HIV in bone mass derangement (Cazanave *et al.*, 2008).

A recent work comparing bone mineral density in 152 HIV-seropositive and 100 healthy women disclosed a highly significant ($p < 0.0001$) decrease in mineralization in HIV subjects, which remained significant also after controlling for race (Dolan *et al.*, 2007). A Canadian Multicentre Study for Osteoporosis reported higher incidences of fractures in HIV patients than in a control group (26.1% vs 17.7%), but failed to find any difference in BMD, suggesting that HIV exerts its influence on bone structure rather than bone mineralization (Prior *et al.*, 2007).

The role of therapy

The true impact of HIV-1 infection and/or HAART therapy in the assessment of bone loss remains largely unsettled (Bongiovanni *et al.* 2003; Klotsas & Klotsas, 2007; Brown & Qaqish, 2006).

The association between osteopenia and PI emerged in several *in vitro* models and seems to have a different etiology depending on specific molecules (Pan *et al.*, 2004, 2006). Malizia *et al.* (2007) performed a microarray analysis on osteoblasts demonstrating a transcription increase in several inflammation related genes, such as

MCP-1 and IL-8, after contact with Nelfinavir, Saquinavir or Ritonavir. Similar results (Jain & Lenhard, 2002) were obtained with Nelfinavir, but not Ritonavir and Saquinavir, able to inhibit both osteoblastic differentiation of mesenchymal stem cells, and decrease osteoblast activity.

Bone metabolism alterations can be determined by functional damage to other organs involved in bone homeostasis such as liver (Collier, 2007) or kidney (Moe et al., 2006). Moreover, PIs can impair vitamin D synthesis *in vitro*, by suppressing the activity of several enzymes involved in its synthesis and catabolism (Cozzolino et al., 2003) inhibiting the osteoblast anabolic activity.

Hence, the control of antiretroviral toxicity and the management of secondary lesions due to both life-long patient treatment and the slower evolution of HIV infection represent a pivotal issue for the clinical and therapeutical approach to HIV positive patients.

Conclusion

Osteopenia and osteoporosis are described in a higher proportion of HIV-infected patients than healthy individuals (Paton et al., 1997; Annapoorna et al., 2004; Glesby, 2003; Amorosa & Tebas, 2006). Although several general mechanisms involved in bone derangement during HIV-1 infection may contribute to decreased bone mineral density in HIV-infected patients, the role

of increased levels of pro-inflammatory cytokines associated with HIV infection, and HIV itself, may also contribute directly to accelerated bone loss (Glesby, 2003; Gibellini et al. 2007). Even though a high incidence of osteopenia/osteoporosis has been associated with both protease inhibitor (PI) and nucleoside/nucleotide reverse transcriptase inhibitor (NRTI)-based treatments (Pan et al., 2006), observations on HIV-1 naïve patients suggested a consistent direct pathogenetic role of HIV-1 (Bruera *et al.* 2003; Gibellini et al. 2007) eliciting the derangement of bone homeostasis. Overall, these studies investigating the relationship between HAART, HIV and osteopenia/osteoporosis show the growing clinical importance of bone damage in the course of HIV infection and the need to disclose the pathogenic mechanisms underlying bone derangement. Hence, the management of bone damage in HIV-infected patients represents a major concern to solve in the near future for a useful follow-up of HIV seropositive patients.

This work was supported by Funds from Fondazione Cassa di Risparmio Bologna, Italy (n° 2006.0035, June 2006), "AIDS projects" (30G.27) of the Italian Ministry of Health, the SIVIM study group for test standardization, funds for selected research topics of the University of Bologna and MURST 60%.

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